

Heterologous expression and characterisation of novel chitinases from a yet unculturable, highly abundant bacterial symbiont of octocorals



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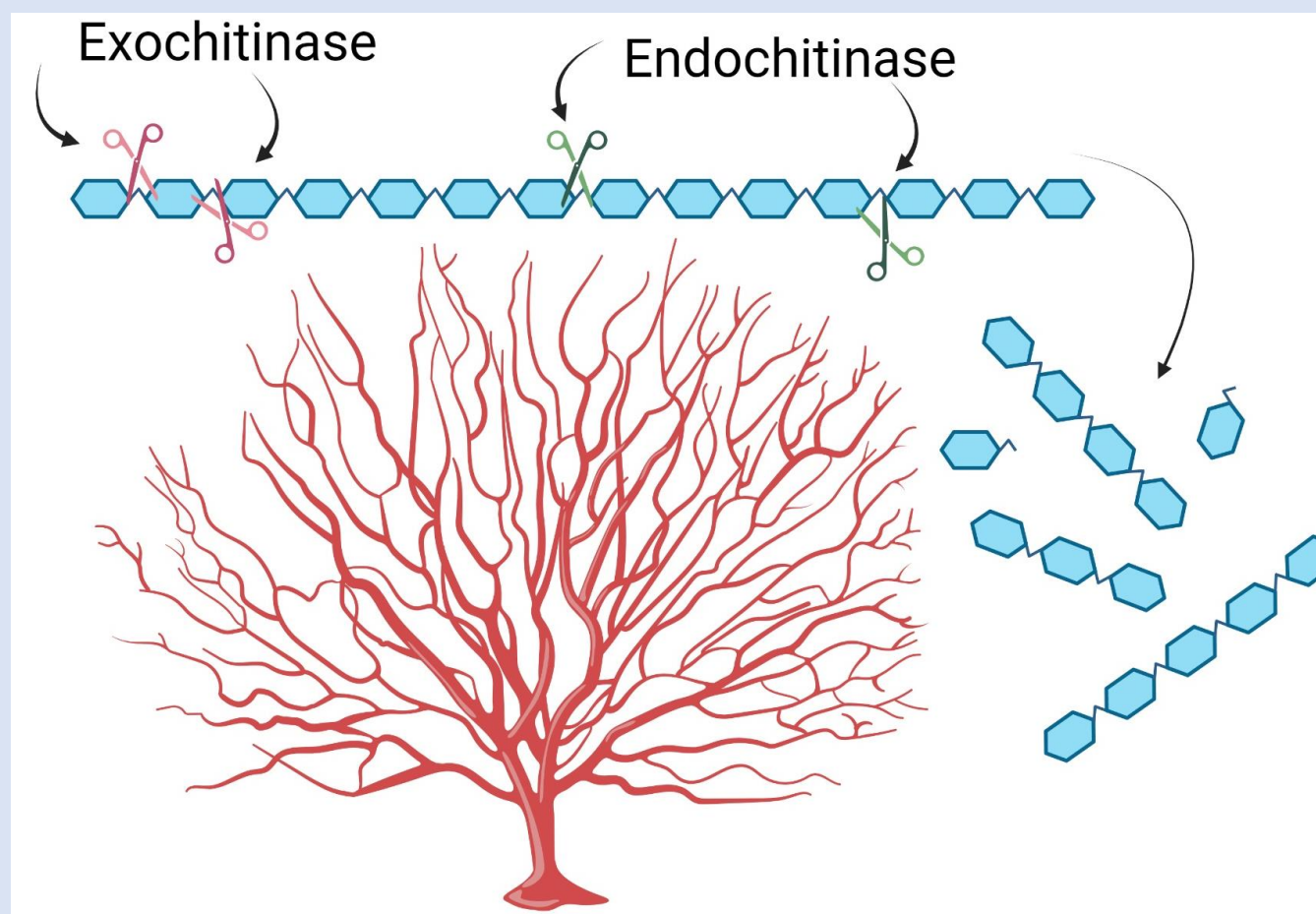
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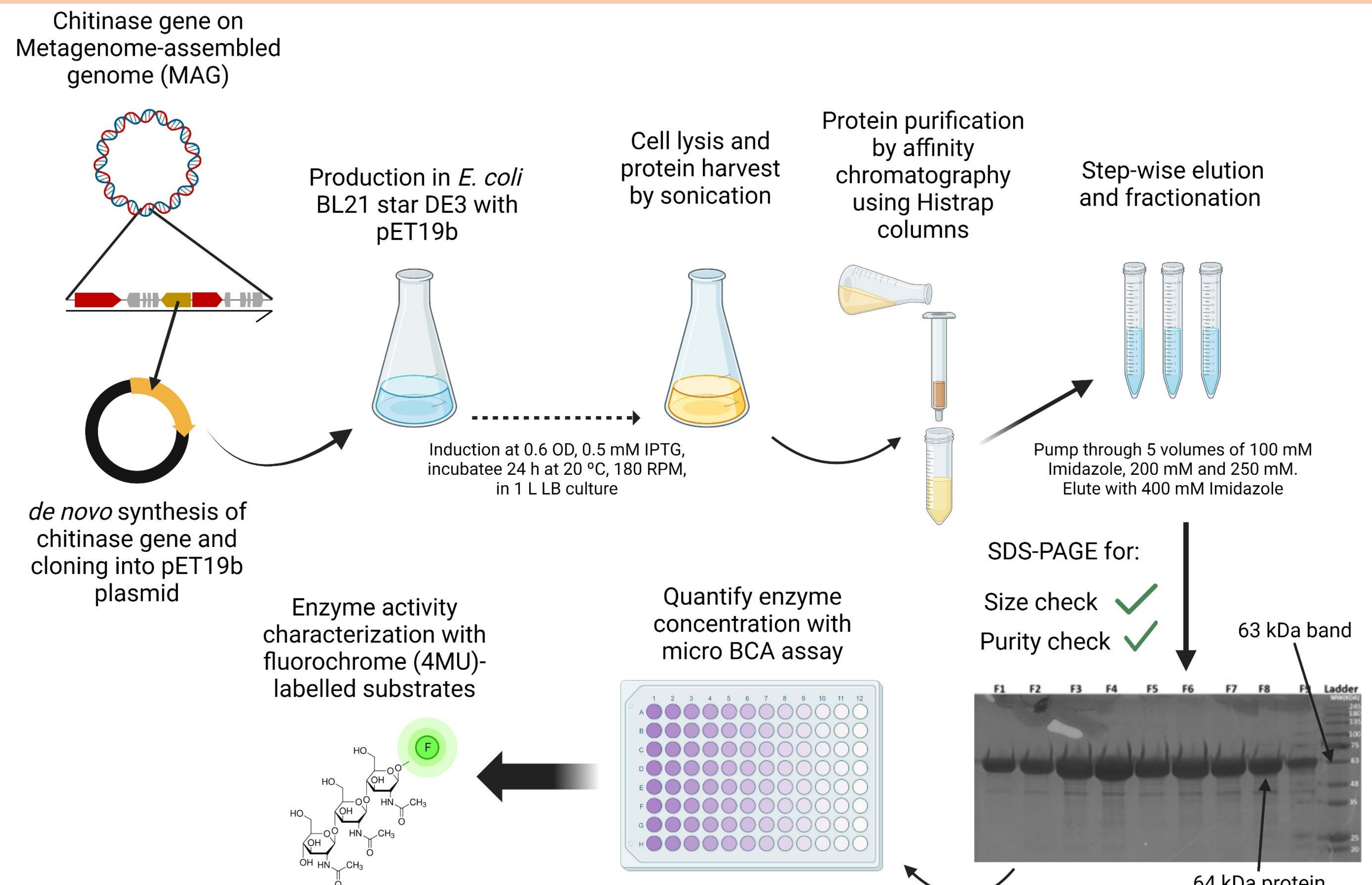
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Introduction

- Chitin is the most abundant natural polymer in marine settings.
- Octocorals (p: Cnidaria) are suspension-feeding marine invertebrates that consume a chitin-rich diet (e.g., zooplankton, diatoms).
- Their microbiomes are likely well adapted to chitinous food processing.
- The marine bacterial family *Endozoicomonadaceae* is an important part of the coral microbiome, reaching high relative abundances and is an indicator of coral health [1].



Methodology: Heterologous chitinase expression and purification



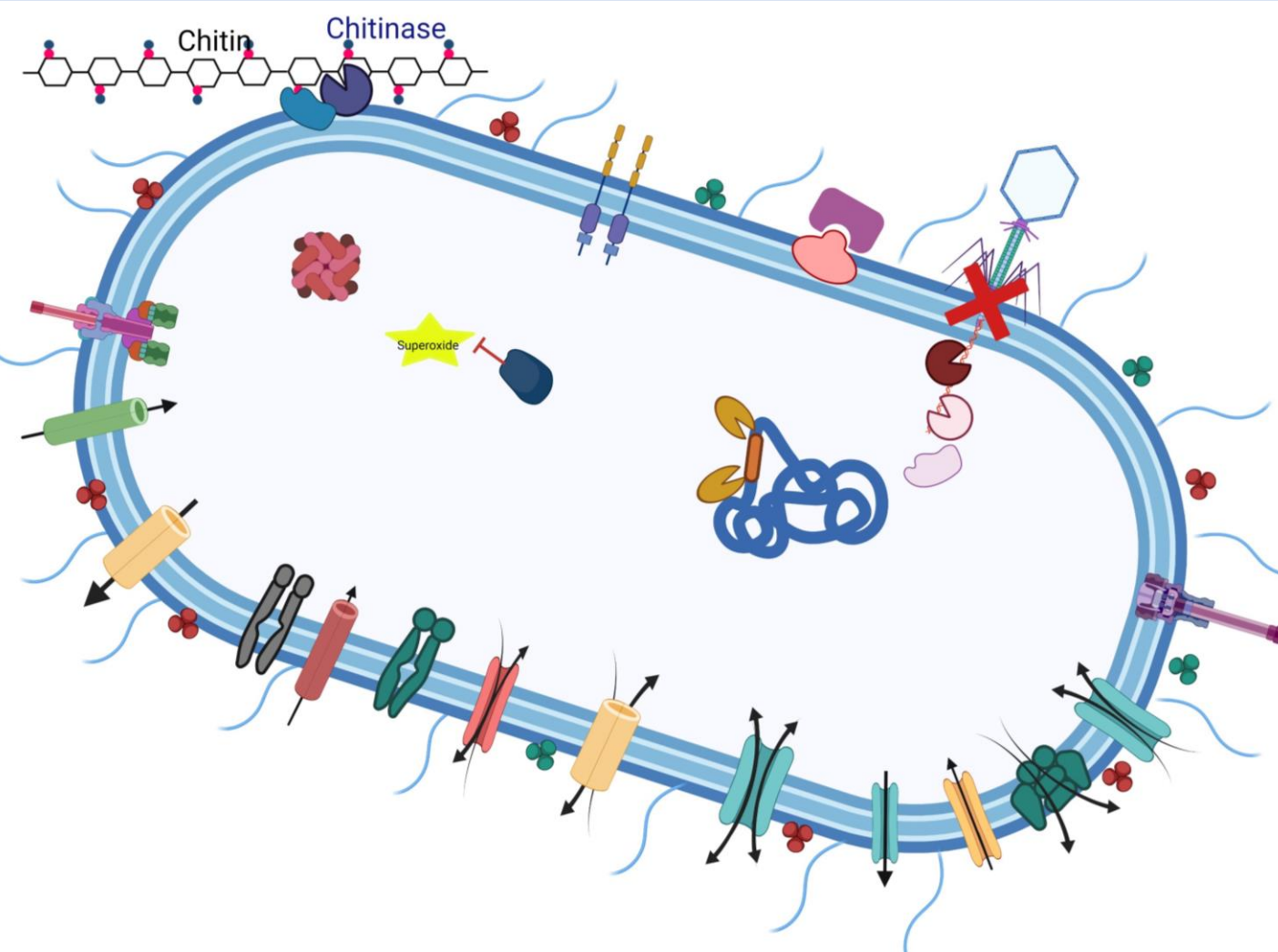
Endozoicomonadaceae symbionts of corals

Involved in amino acid and vitamin B supply to the host, micronutrient acquisition, sulfur cycling and carbohydrate metabolism [2].

Only a few species are successfully cultured and maintained in the laboratory, severely limiting investigation of their ecology & physiology.

A novel, yet uncultured genus, *Candidatus Gorgonimonas*, from the Endozoicomonadaceae family was proposed by our team in 2022 [2].

A metagenomics survey showed that *Candidatus Gorgonimonas* harbours endo-chitinase genes with low homology to known chitinases (< 50%) [3].

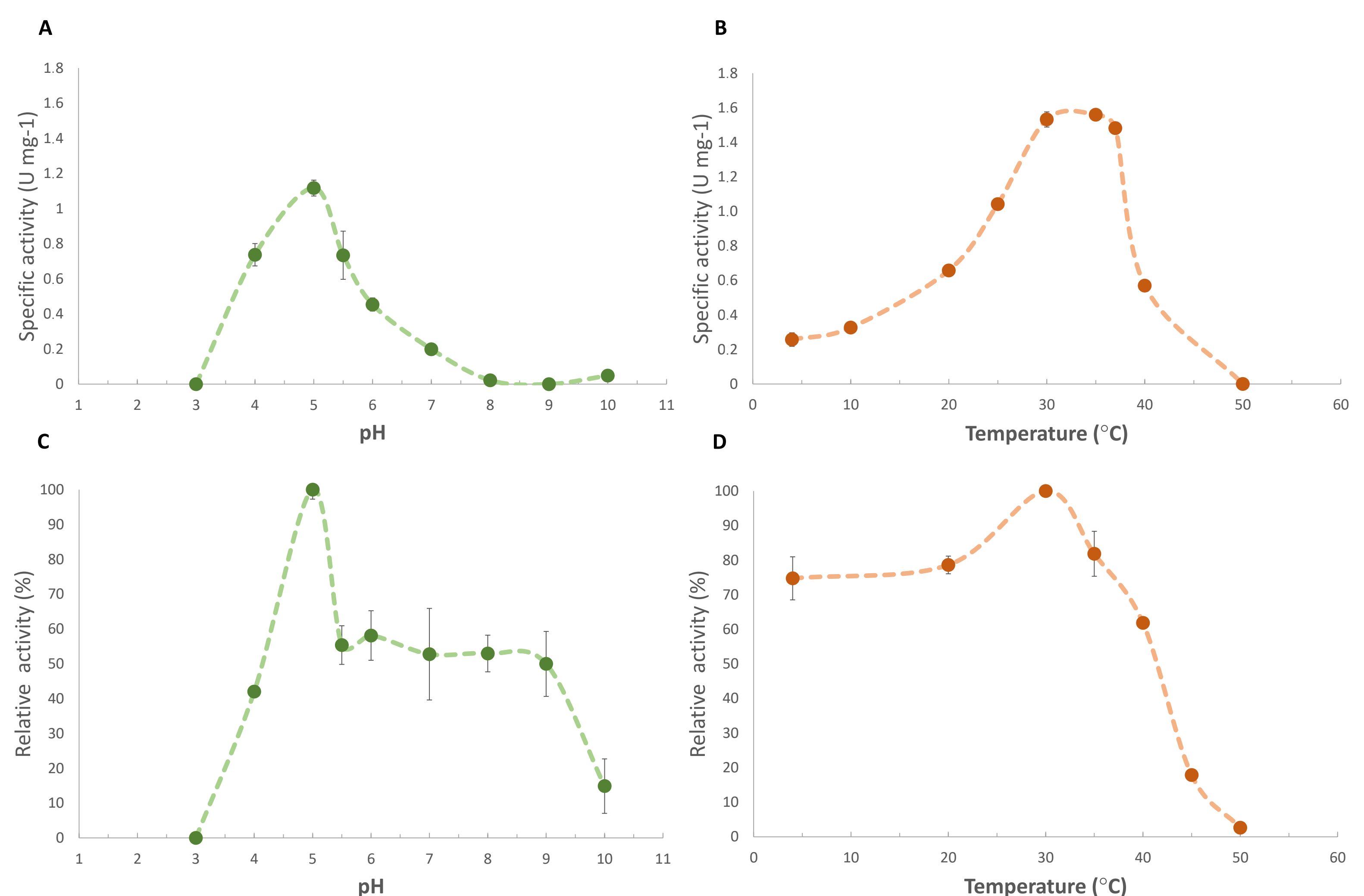


[1] Keller-Costa et al., 2021, Microbiome, 9, 1-21 <https://doi.org/10.1186/s40168-021-01031-y>

[2] Keller-Costa et al., 2022, Microbiome 10, 151 <https://doi.org/10.1186/s40168-022-01343-7>

[3] Silva et al., 2023, ISME Communications 3, 109 <https://doi.org/10.1038/s43705-023-00316-7>

Temperature- and pH-dependent activity and stability of the chitinase



Objectives

- Synthesize *de novo* the chitinase gene from uncultured octocoral symbiont *Candidatus Gorgonimonas* and express it in *E. coli*.
- Characterize the chitinase activity over a range of temperatures and pH with fluorochrome-labeled substrates.
- Analyze the substrate consumption (colloidal chitin and chitosan) and chitin oligomer production pattern of the enzyme (Ongoing!).

Metagenomics-based predictions of the chitinase:

The native gene was 1644 bp, and the native protein is 576 aa (pI 4.89, 64.6 KDa).

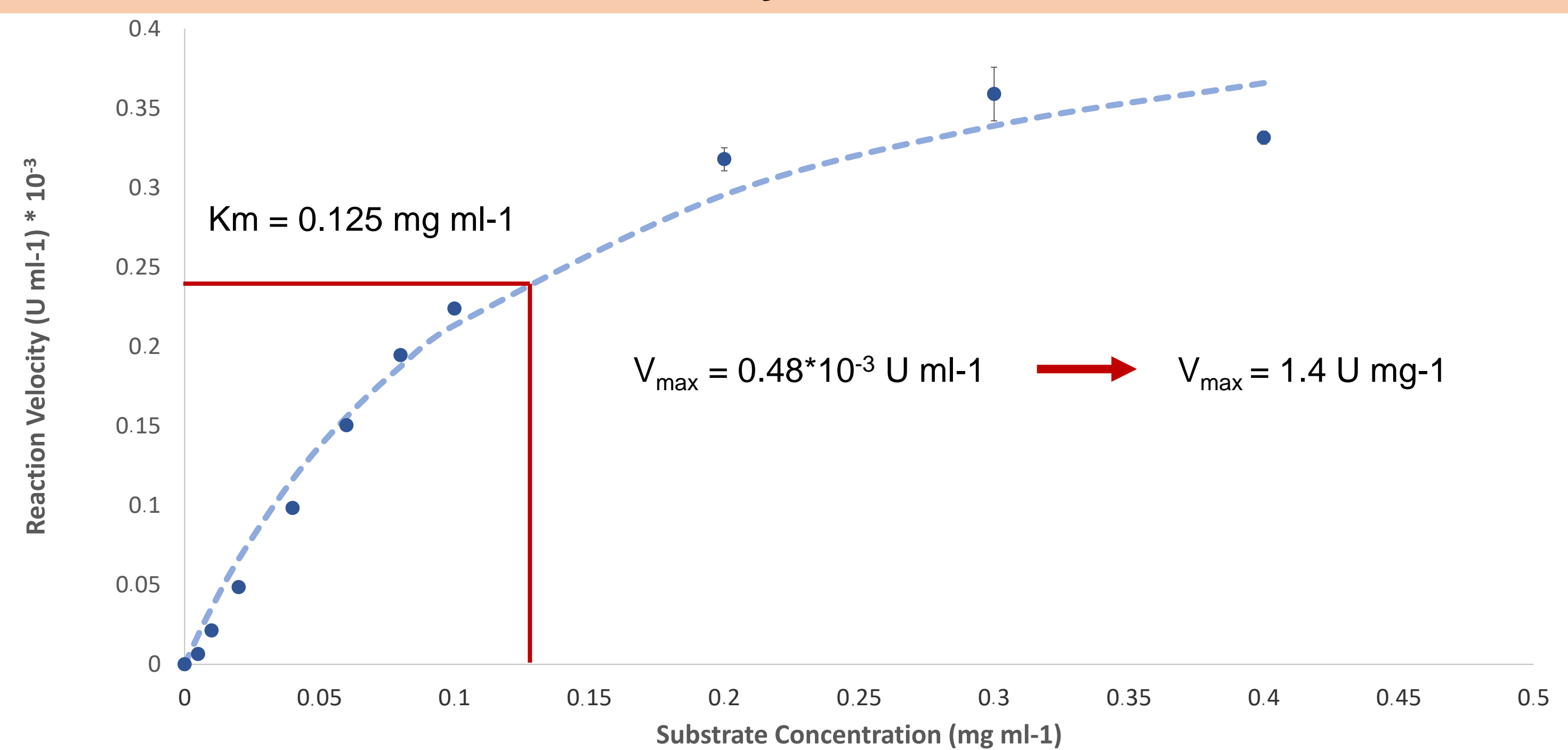
The recombinant protein (- signal peptide, + Histidine tail) is 571 aa, has a pI of 5.13 and 64.1 Kda.

The enzyme was predicted to be an endochitinase (EC 3.2.1.14), it had a GH18 protein family domain, and two unknown domains, DUF5011 and DUF5011.

Conclusion

- The here produced recombinant protein from yet uncultured octocoral symbiont *Candidatus Gorgonimonas* presents endo-chitinase activity but no N-acetylglucosaminidase (exo-chitinase) activity.
- The endochitinase was active in the range of 4 – 40 °C, and 4 – 7 pH, with highest activity (1.6 U mg⁻¹) being reached at 30 – 37 °C and pH 5.
- Although stability is highest at pH 5, the chitinase is also stable at neutral to alkaline pH (until pH 9) values and temperatures up to 40 °C, retaining > 50 % activity.
- The chitinase presents a V_{max} of $0.48 \cdot 10^{-3}$ U ml⁻¹ and a K_m -value of 0.125 mg ml⁻¹.
- Releases chitin trimers (triacylchitotriose) and dimers (diacylchitobioside), but not monomers (N-acetylglucosamine).

Kinetic characterization with 4-Methylumbelliferyl β -DN,N',N''-triacylchitotriose



Kinetic characterization was conducted using Citric acid – Trisodium citrate 10 mM as buffer at pH 5, at 35 °C.